FIRST RECORD OF *Molossus coibensis* (CHIROPTERA: MOLOSSIDAE) IN THE BRAZILIAN AMAZON

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ABSTRACT. This study reports on the occurrence of *Molossus coibensis* J. A. Allen, 1904, in the Brazilian Amazon and the second record of this species from Brazil. It also presents a comparative analysis with other two small-bodied *Molossus* species that occur in Brazil: *M. molossus* (Pallas, 1766) and *M. aztecus* Saussure, 1860. The report is based on 8 females captured in urban areas of the city of Bragança, in the northern Brazilian state of Pará. Morphometric data and the qualitative comparison of morphological features permitted the identification of the specimens as *M. coibensis*, hitherto known only in Tapirapuã, in the southwestern state of Mato Grosso, west-central Brazil.


Key words: Coiban mastiff bat. New record. Northern Brazil.


The bats of the genus *Molossus* Geoffroy, 1805 have an essentially Neotropical distribution, with a total of 7 or 8 species, depending on the taxonomic arrangement (Simmons, 2005; Eger, 2007). Six species are known to occur in Brazil: *M. coibensis* J. A. Allen, 1904; *M. currentium* Thomas, 1901; *M. molossus* (Pallas, 1766); *M. pretiosus* Miller, 1902; *M. rufus* E. Geoffroy, 1805, and *M. aztecus* Saussure, 1860, which was recently recorded in southern Minas Gerais state (Gregorin et al., 2011). Of these species, *M. currentium* has been recorded in Manaus,
in the state of Amazonas (López-Gonzáles and Presley, 2001), although this record is based on a juvenile specimen which was considered uncertain by Eger (2007). Prior to the present study, the only record of *M. coibensis* from Brazil was a specimen collected at Tapirapoan, in the state of Mato Grosso, which consists only of a preserved skin, given that the skull has been lost (Dolan, 1989; Eger, 2007).

Coiban Mastiff Bat, *M. coibensis* (Fig. 1), was first described from the Island of Coiba in Panama by Allen (1904). The known geographic distribution of this species (Fig. 2) includes the whole of Central America, in particular the Pacific coast, extending between Chiapas (Mexico) and northern South America (Ecuador, Colombia, Venezuela, Peru, and Guiana), it is noteworthy that French Guiana is the type locality of *Molossus barnesi*, considered by some as synonymous of *M. coibensis* (Eger, 2007). Its distribution also extends to southwestern Brazil (Dolan, 1989). Paglia et al. (2012) included the geographic range of *M. coibensis* in both the Brazilian Amazon and Cerrado biomes, although the only Brazilian locality for this species, Tapirapoan (14°85’ S, 57°75’ W), is located in the municipality of Barra do Bugres in southwestern Mato Grosso, within the Cerrado savanna, but close to its limits with the Amazon and Pantanal biomes (Paglia et al., 2012: fig. 6).

As the diversity of *Molossus* in South America is still poorly understood and the sampling of some species is scarce, the present record of *M. coibensis* was confirmed through comparisons with other specimens of the same species from Central America, and other small-bodied species, which confirmed the identity of the specimens and the occurrence of *M. coibensis* in northern Brazil.

Fig. 1. Adult female *Molossus coibensis* captured in the town of Bragança, in the northern Brazilian state of Pará.

Six adult and two juvenile female individuals of *M. coibensis* were collected during an inventory of the bat fauna at Bragança, in the Brazilian state of Para (01°03’13” S, 46°45’56” W), carried out between August, 2007, and August, 2008. The bats were captured in the roofs of houses covered with cement-fiber tiles in the center of the town. Each specimen was measured (external dimensions and cranial/dental features) and weighed, and its age and sex were determined. Biometric parameters were based on those analyzed by Dolan (1989) and Gregorin et al. (2011) – total length of the cranium (TLC), condylo-incisor length (CIL), upper tooth row length, from the canine to the last molar (C-M), brain case breadth (BCW),
breadth of the post-orbital constriction (POC), maximum breadth across the upper molars (M-M), maximum breadth across the upper canines (C-C). The specimens were fixed in 10% formalin and preserved in 70% ethanol prior to being deposited in the Coleção de Zoologia da Universidade Federal do Pará Campus de Bragança (LJCC – field number) and the Coleção de Mammíferos da Universidade Federal de Lavras em Minas Gerais (CMUFLA). For the comparative analysis, specimens were examined in the collections of the American Museum of Natural History (AMNH) in New York, the US Natural History Museum (USNM) at the Smithsonian Institution in Washington D.C., The Field Museum (FMNH) in Chicago, and the Museum of Comparative Zoology (MCZ) at Harvard University (Appendix).

Given the marked sexual dimorphism in *Molossus* (Dolan, 1989) and the fact that all the specimens collected in the present study were females, morphometric analyses included only adult females. The only male specimens examined in this study were the holotypes of *M. cherriei* (= *M. coibensis*) and *M. coibensis*, which were included in the analysis of qualitative variables.

The *M. coibensis* specimens presented here were identified based on the diagnosis of Dolan (1989) and comparisons with specimens from Panama, close to the type locality (Coiba Island). The diagnostic characteristics of the specimens were (1) small forearm (Table 1), (2) short cranium with more convex brain case (Table 1; Figs. 3 and 4), (3) upper incisors short and convergent or blade-like (Fig. 3),

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### Table 1

Morphometric parameters for adult female of *Molossus coibensis* (northern Brazil and Panama), *M. aztecus* (Mexico), and *M. molossus* (northern Brazil). The variables are described in the text. Mean ± standard deviation in millimeters, range of values and sample size (n).

<table>
<thead>
<tr>
<th>Parameter</th>
<th><em>M. coibensis</em> (Bragança, Brazil)</th>
<th><em>M. coibensis</em> (Panama)</th>
<th><em>M. aztecus</em> (Mexico)</th>
<th><em>M. molossus</em> (Para, Brazil)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Length of forearm</td>
<td>34.3 ± 0.4 (33.5-34.7)</td>
<td>34.9 ± 1.3 (32.8-37.5)</td>
<td>36.1 ± 0.5 (35.5-36.7)</td>
<td>40.9 ± 1.3 (38.5-43.6)</td>
</tr>
<tr>
<td>Cranium length</td>
<td>15.8 ± 0.2 (15.5-16.1)</td>
<td>15.8 ± 0.6 (15.1-17.2)</td>
<td>16.5 ± 0.1 (16.3-16.6)</td>
<td>16.7 ± 0.3 (16.2-17.6)</td>
</tr>
<tr>
<td>Condylo-incisor length</td>
<td>14.6 ± 0.3 (14.3-15.1)</td>
<td>14.8 ± 0.5 (14.0-15.8)</td>
<td>15.3 ± 0.2 (15.0-15.6)</td>
<td>15.9 ± 0.5 (14.3-16.8)</td>
</tr>
<tr>
<td>Length of upper tooth row</td>
<td>5.9 ± 0.2 (5.7-6.1)</td>
<td>5.9 ± 0.2 (5.7-6.2)</td>
<td>6.2 ± 0.1 (6.1-6.4)</td>
<td>6.3 ± 0.1 (6.1-6.6)</td>
</tr>
<tr>
<td>Brain case breadth</td>
<td>8.8 ± 0.2 (8.6-9.0)</td>
<td>8.7 ± 0.2 (8.3-9.2)</td>
<td>8.8 ± 0.2 (8.7-9.1)</td>
<td>9.0 ± 0.4 (8.5-10.7)</td>
</tr>
<tr>
<td>Maximum breadth between upper molars</td>
<td>7.5 ± 0.2 (7.2-7.8)</td>
<td>7.5 ± 0.3 (7.2-8.1)</td>
<td>7.9 ± 0.3 (7.6-8.4)</td>
<td>7.8 ± 0.3 (7.4-9.1)</td>
</tr>
<tr>
<td>Breadth of the post-orbital constriction</td>
<td>3.9 ± 0.1 (3.7-4.0)</td>
<td>3.6 ± 0.1 (3.4-3.9)</td>
<td>3.7 ± 0.1 (3.7-3.8)</td>
<td>3.7 ± 0.2 (3.4-4.1)</td>
</tr>
<tr>
<td>Maximum breadth across the upper canines</td>
<td>4.6 ± 0.2 (4.3-5.0)</td>
<td>4.1 ± 0.1 (3.9-4.4)</td>
<td>4.5 ± 0.1 (4.4-4.5)</td>
<td>4.1 ± 0.1 (4.1-4.2)</td>
</tr>
</tbody>
</table>

* Data compiled from Dolan (1989);
** Data compiled from Dolan (1989) for the southernmost locality analyzed by this author (El Salvador).
(4) infraorbital foramen opening more laterally than frontally (Figs. 3 and 4; see Gregorin et al., 2011), (5) blackish pelage, with short (2.3-3.7 mm) dorsal hairs around the shoulders, and (6) hairs with grayish basal third, resulting in a weak contrast with the apical band (Dolan, 1989).

Of these characters, 2 and 6 are shared by *M. coibensis*, *M. aztecus*, *M. pretiosus*, and *M. rufus*, and distinguish these species clearly from *M. molossus*. In fact, the pelage of *M. molossus* is typically chestnut, with relatively long hairs characterized by a white basal band which contrasts distinctly with the apical band; the rostrum in *M. molossus* is narrower, with a proportionately narrow brain case (Table 1); the upper incisors are elongated with parallel (pincer-like) tips (Fig. 1), distinct of blade-like upper incisors of *M. coibensis*. In comparison with *M. aztecus*, a closely-related small-bodied species, *M. coibensis* is differentiated by much smaller external measurements and cranial/dental parameters (Table 1).

The occurrence of *M. coibensis* in the Brazilian Amazon basin would be expected from its distribution in northern South America, including the Guianas, Colombia, Venezuela, and Peru (Eger, 2007), although the only previous record from Brazil, at Tapirapoan in Mato Grosso, was from the Cerrado savanna. The only Brazilian specimen available prior to the present study (AMNH 36699) now consists only of the skin, given that the skull was lost. This specimen is the holotype of *M. cherriei*, described by Allen (1916), which was subsequently synonymized with *M. coibensis* by Dolan (1989). The characteristics of this specimen, which include a short forearm (33.8 mm),
gray-blackish pelage, and short dorsal hairs with a grayish base and weak banding, are clearly consistent with the diagnostic traits of *M. coibensis*.

The occurrence of *M. coibensis* at such widely separated localities in Brazil indicates that this species is likely also found throughout much of the Brazilian Amazon basin, and possibly also in the Pantanal wetlands. The paucity of records of this species within Brazilian territory is probably due to the general lack of data on molossids in most regions, and the difficulties in identifying the species of this group.

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**LITERATURE CITED**


**APPENDIX**

**Examined material**